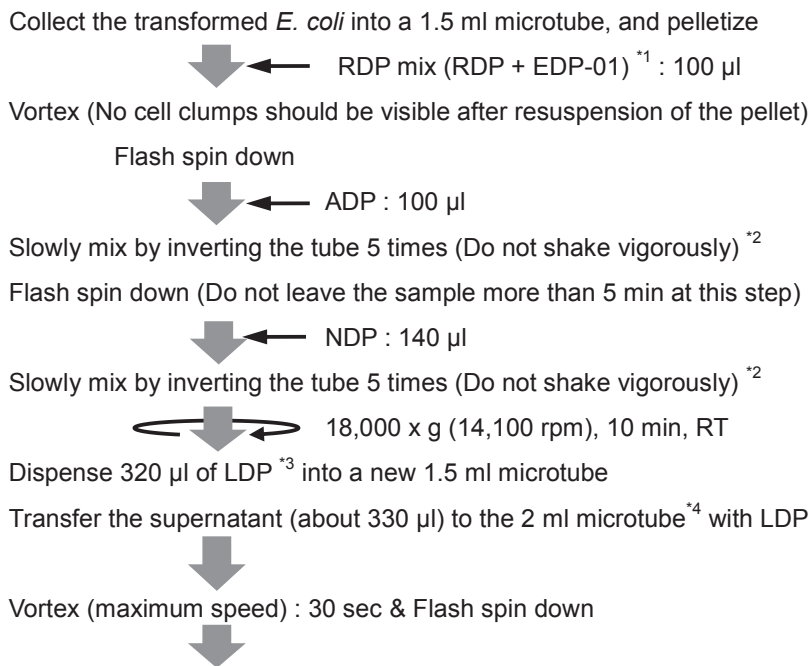


Automated Plasmid DNA Extraction from *E. coli*

Protocol



Set into the device
Protocol: PLASMID
(Elution volume : 50 μ l ^{*5})

*Please refer to Quick Start Guide or operation manual to know how to set sample tube.

1. Apply the lysate into the cartridge
2. Pressurizing
3. Wash 2 times by Wash Buffer (WRT)
4. Add selected volume of Elution buffer and elute plasmid DNA into collection tube.

Plasmid DNA

^{*1} Before starting an extraction experiment, add total amounts of EDP-01 to RDP bottle, and mix well. In the case of storing RDP mix, it is recommended to preserve it under refrigeration (2-8°C) and use within 6 months.

^{*2} After addition of ADP or NDP, immediately mix by inverting the tube 5 times. Vigorous mixing results in the co-purification of much of genomic DNA. Too slow mixing causes inadequate blending of liquids, resulting in deterioration in the yield of plasmid DNA.

^{*3} Add 44 ml of >99% ethanol into the bottle and mix well by gently inverting the bottle before use.

^{*4} Following microtube are recommended.
#BM4020 (BM instrument co., ltd)
#72.695.700,
#72.695.500S (SARSTEDT)

^{*5} The volume of the eluate from each cartridge is 100 μ l. The volume of CRT can be reduced to 50 μ l, but in that case, elution efficiency might be decreased.

Results

The yield of plasmid DNA / Protein contamination : A260/280
/ Chaotropic salt contamination : A260/230

Kit	Yield	A260/280	A260/230
QuickGene	29.5 µg	1.98	2.23

N=4

Common protocol is usable for the following

Fosmid

Contact Information

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Depending on sample and storage conditions, nucleic acid may not be extractable.
Therefore, we cannot guarantee accurate data.
The extracted nucleic acid contains unintended acid (ex: when extracting DNA, RNA is also extracted).

