



QuickGene-AutoS Plasmd kit (AS-PL)

## Automated Plasmid DNA Extraction from *E. coli*

## Protocol

Collect the transformed E. coli into a 1.5 ml microtube, and pelletize

■ RDP mix (RDP + EDP-01) \*1 : 100 µI

Vortex (No cell clumps should be visible after resuspension of the pellet) Flash spin down

**←** ADP : 100 μl

Slowly mix by inverting the tube 5 times (Do not shake vigorously) \*2

Flash spin down (Do not leave the sample more than 5 min at this step)

✓ NDP : 140 µl

Slowly mix by inverting the tube 5 times (Do not shake vigorously) \*2

18,000 x g (14,100 rpm), 10 min, RT

Dispense 320 µl of LDP \*3 into a new 1.5 ml microtube

Transfer the supernatant (about 330 µl) to the 2 ml microtube 4 with LDP



Vortex (maximum speed): 30 sec & Flash spin down

Set into the device

Protocol: PLASMID (Elution volume : 50 µl \*5)

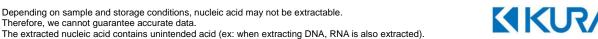
> \*Please refer to Quick Start Guide or operation manual to know how to set sample tube.



- 1. Apply the lysate into the cartridge
- 2. Pressurizing
- Wash 2 times by Wash Buffer (WRT)
- Add selected volume of Elution buffer and elute plasmid DNA into collection tube.

Plasmid DNA

- \*1 Before starting an extraction experiment, add total amounts of EDP-01 to RDP bottle, and mix well. In the case of storing RDP mix, it is recommended to preserve it under refrigeration (2-8°C) and use within 6 months.
- \*2 After addition of ADP or NDP, immediately mix by inverting the tube 5 times. Vigorous mixing results in the co-purification of much of genomic DNA. Too slow mixing causes inadequate blending of liquids, resulting in deterioration in the yield of plasmid DNA.
- \*3 Add 44 ml of >99% ethanol into the bottle and mix well by gently inverting the bottle before use.
- \*4 Following microtube are recommended. #BM4020 (BM instrument co., Itd) #72.695.700, #72.695.500S (SARSTEDT)
- \*5 The volume of the eluate from each cartridge is 100 μl. The volume of CRT can be reduced to 50 µl, but in that case, elution efficiency might be decreased.







## Results

The yield of plasmid DNA / Protein contamination: A260/280 / Chaotropic salt contamination: A260/230

Kit	Yield	A260/280	A260/230
QuickGene	29.5 μg	1.98	2.23

N=4

# Common protocol is usable for the following

Fosmid

## **Contact Information**

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Depending on sample and storage conditions, nucleic acid may not be extractable. Therefore, we cannot guarantee accurate data.

The extracted nucleic acid contains unintended acid (ex: when extracting DNA, RNA is also extracted).





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